ELSEVIER

Contents lists available at ScienceDirect

Bioorganic & Medicinal Chemistry Letters

journal homepage: www.elsevier.com/locate/bmcl



Crown-capped imidacloprid: A novel design and insecticidal activity

Shinzo Kagabu ^{a,*}, Masaru Takagi ^a, Ikuya Ohno ^a, Tsuyoshi Mikawa ^b, Toru Miyamoto ^b

ARTICLE INFO

Article history:
Received 10 March 2009
Revised 27 March 2009
Accepted 15 April 2009
Available online 18 April 2009

Keywords: Crown-capped insecticide Imidacloprid Neonicotinoid insecticides

ABSTRACT

Imidacloprid (IMI) derivatives conjugated with benzo-15-crown-5 and benzo-18-crown-6 structures, applied for the first time to explore novel insecticidal molecule, elicited strong excitatory toxic signs to the house flies and stunningly exhibited three to five times higher insecticidal activity than that of the parent IMI, yet the two benzo-crown structures themselves had no effect.

© 2009 Elsevier Ltd. All rights reserved.

The use of crown ethers expanded in analytical chemistry, organic synthesis, biological systems, molecular electronics, and other material technologies, resulting in an increasing number of recent breakthroughs. However, to our knowledge, pesticides designed using supramolecular aspects have not been published. The present investigation applies for the first time the crown-ether structure to design novel insecticidal molecules, leading to benzo-15-crown-5 or benzo-18-crown-6 conjugated imidacloprid (IMI). IMI is the current top-selling insecticide acting at the insect nicotinic receptor. The prosthetic crown-ether moiety with a specific Na⁺ carrier and amphiphilic functions may give unexpected biological properties. Intriguingly, we found that the crown-capped IMI derivatives showed high insecticidal activity consistent to or greater than that of the parent IMI.

Benzo-15-crown-5 and benzo-18-crown-6 capped imidacloprids (**2**, **3**) were prepared by the reaction of 2-nitroiminoimidazolidine-1-ylmethyl-benzo-crown (**7**, **8**) with 6-chloro-3-pyridylmethyl chloride and resulted in yields of 33% and 45%, respectively (Fig. 1).⁴

A test compound (0.22 μ L, dissolved in 80% DMSO) was injected into the thorax of female adult house flies (Musca~domestica). Mortality was determined at 24 h after treatment. The LD₅₀ values were calculated using log dose-probit mortality analysis. The toxicity signs of the poisoned flies were observed in the following sequence: convulsions with leg tremor and wing motion followed by paralysis, and death with a curled abdomen or out-stretched legs, similar to that of IMI itself, suggesting that the crowned derivatives presumably act on the insect nervous system. The LD₅₀s were 25 \pm 4.4 and 17 \pm 2.4 ng/fly (n = 5) for 2 and 3, respectively. Under these conditions, the LD₅₀ for IMI was 84 \pm 5.1 ng/fly (n = 3). Inter-

estingly, neither of benzo-15-crown-5 nor benzo-18-crown-6 showed toxic effect even at doses as high as 1100 ng/fly. Moreover, the insecticidal activities of the crowned compounds were determined in the presence of the metabolic inhibitor, propyl 2-propynyl phenylphosphonate known to synergize the insecticidal potency of IMI.⁶ Insecticidal potency of compounds 2 and 3 in the presence of the metabolic inhibitor were 4.8 ± 1.8 and 4.3 ± 1.1 ng/fly (n = 3), respectively, which were higher than that of IMI. When the compounds topically-applied to the insect integument at a dose of 15 μ g/fly, the mortality was 44% and 38% for 2 and 3, respectively. These values were higher than the 24% observed for IMI. Lipophilic ingredients are generally more effective upon insects in integument penetration and distribution in the body, which results in efficient delivery to the site of action through the lipid-protein membrane. Several experiments using IMI-related compounds have actually shown that the higher the hydrophobicity of the molecules, the higher the insecticidal or neurophysiological activity.8

One of the most prevalent hydrophilicity/lipophilicity quotient is the partition coefficient between octanol and water (Pow). The log Pow for $\bf 2$ and $\bf 3$ were 0.15 and 0.28 (n = 4), respectively. These values were smaller than observed 0.57 for IMI. This suggested that crowned derivatives are more hydrophilic than IMI. The unexpectedly higher dermal insecticidal potencies of the crowned derivatives despite their smaller Pows may be explained by the amphiphilic properties of crown ether. Crown ether can be lipophilic or hydrophilic depending on the ring conformation, which can change according to the properties of the surrounding fluid (Fig. 2).

We showed that crown-capped IMI derivatives have higher insecticidal activity compared with the parent IMI. Considering that molecules having potencies comparable to the most active product in this insecticide class have rarely been found despite the number

a Department of Chemistry, Faculty of Education, Gifu University, Gifu 501-1193, Japan

^b Department of Applied Biology and Chemistry, Tokyo University of Agriculture, Tokyo 156-8502, Japan

^{*} Corresponding author. Tel.: +81 58 293 2253; fax: +81 58 293 2207. E-mail address: kagabus@gifu-u.ac.jp (S. Kagabu).

Figure 1. limidacloprid (IMI) and preparation of crowned derivatives.

 $\textbf{Figure 2.} \ \ \text{Conformational change of benzocrowns.} \ \ \textbf{I} \ \ \text{and} \ \ \textbf{II} \ \ \text{in lipophilic and hydrophilic fluids, respectively.}$

of structural modifications testing various angles that have been attempted over the past 30 years, it was surprising that the activity of IMI was enhanced by crown-bridging to the extent we observed. ¹⁰ Therefore we questioned whether the crowned-IMI was the real active species at the target site, or if we were observing the effect of a physiologically regenerated IMI. Our experiments showed that no detectable amount of liberated IMI were observed after 24 h in a physiological salt solution. This result caused us to exclude the effect of a spontaneous prodrug from our observations. ¹¹

To approach the effect of crown-capping on the biological action of IMI, we examined the complexing behavior of the derivatives in the presence of several molar ratios of sodium thiocyanate by NMR in CDCl₃/CD₃OD (1:50 v/v) according to the standard protocol.¹² Both insecticidal molecules showed a typical profile representing a loosely bound 1:1 complex with Na[†]. The binding constants were as high as 440 ± 30 and $16,000 \pm 3000$, respectively, at 25 °C (n = 3) for **2** and **3**, comparable to those of the uncapped benzo-crown compounds.¹³ Therefore, the crown-capped derivatives have prominent ion-capturing properties, conceivably attenuating physiology of the nervous system and ultimately conferring the enhanced insecticidal potency.

In conclusion, the novel crown-capped IMI derivatives had high insecticidal potency, suggesting an action at the insect nervous system presumably mediated by nicotinic receptor. The biochemical and physiological studies should be conducted to define the unique mode of action for the crown-capped insecticide.

Acknowledgments

We thank Professor M. Kawasaki and Dr. C. Ikeda for their helpful comments.

References and notes

- Vögtle, F. Supramolecular Chemistry: An Introduction; Wiley: Chichester, UK, 1993; Vol. 5; Lehn, J.-M. Supramolecular Chemistry: Concepts and Perspectives; VCH: Weinheim, Germany, 1995; Steed, J. W.; Atwood, J. L. Supramolecular Chemistry; Wiley: Chichester, UK, 2000; Analytical Methods in Supramolecular Chemistry; Schalley, C., Ed.; Wiley-VCH: Weinheim, Germany, 2008; Modern Supramolecular Chemistry: Strategies for Macrocyclic Synthesis; Diederich, F., Stang, P. J., Tykwinski, R. R., Eds.; Wiley-VCH: Weinheim, Germany, 2008.
- The Pesticide Manual; Tomlin, C. D., Ed., 14th ed.; British Crop Protection Council: Hampshire, UK, 2006.
- 3. Moffat, A. S. Science 1993, 261, 550; Kagabu, S. Rev. Toxicol. 1997, 1, 75; Nicotinoid Insecticides and the Nicotinic Acetylcholine Receptor; Yamamoto, I., Casida, J. E., Eds.; Springer: Tokyo, Japan, 1999; Tomizawa, M.; Casida, J. E. Annu. Rev. Pharmacol. Toxicol. 2003, 48, 339; Ihara, M.; Okajima, T.; Yamashita, A.; Oda, T.; Hirata, K.; Nishiwaki, H.; Morimoto, T.; Akamatsu, M.; Ashikawa, Y.; Kuroda, S.; Mega, R.; Kuramitsu, S.; Sattelle, D. B.; Matsuda, K. Invert. Neurosci. 2008, 8, 71; Jeschke, P.; Nauen, R. Pest Manag, Sci. 2008, 64, 1084.
- **2008**, 8, 71; Jeschke, P.; Nauen, R. *Pest Manag. Sci.* **2008**, *64*, 1084. Compound **2**: mp 27–29 °C (from ether). ¹H NMR δ (CDCl₃): 3.50 (4H, m, NCH_2CH_2N), 3.76 (8H, m, $OCH_2CH_2OCH_2CH_2O$), 3.91 (4H, m, 2 × PhOCH₂CH₂), 4.13 (4H, 2 × PhOCH₂CH₂), 4.40 (2H, s, Py-CH₂), 4.48 (2H, s, Ph-CH₂), 6.82 (3H, m, PhH), 7.36 (1H, d, J = 8.2 Hz), 7.72 (1H, dd, J = 8.2/2.6 Hz), 8.31 (1H, d, J = 2.6 Hz). ¹³C NMR δ (CDCl₃): 44.59, 45.11, 47.53, 50.61, 69.04, 69.096, 69.51, 69.55, 70.49, 71.14, 113.86, 113.98, 121.80, 125.01, 126.46, 128.97, 139.27, 149.48, 149.57, 149.78, 152.02, 161.22. FABHRMS for C₂₄H₃₀ClN₅O₇: Calcd 536.1912. Found: 536.1934. Compound **3**: mp 33–36 °C (from ether). 1 H NMR δ (CDCl₃): 3.50 (4H, m, NCH₂CH₂N), 3.68 (4H, br s, OCH₂CH₂O), 3.71 (4H, m, OCH₂CH₂O), 3.77 (4H, m, OCH₂CH₂O), 3.92 (4H, m, 2 × PhOCH₂CH₂), 4.14 (4H, 2 × PhOCH₂CH₂), 4.40 (2H, s, Py-CH₂), 4.48 (2H, s, Ph-CH₂), 6.81 (3H, m, PhH), 7.37 (1H, d, J = 8.2 Hz), 7.73 (1H, dd, J = 8.2/2.6 Hz), 8.31 (1H, d, J = 2.6 Hz). ¹³C NMR δ (CDCl₃): 44.60, 45.08, 47.54, 50.61, 69.17, 69.22, 69.61, 69.65, 70.78, 70.94, 113.91, 114.12, 121.81, 125.05, 126.43, 128.96, 139.29, 149.42, 149.48, 149.62, 152.06, 161.22. FABHRMS for C₂₆H₃₄ClN₅O₈: Calcd 580.2174. Found: 580.2173.
- 5. The houseflies in all experiments were susceptible Takatsuki (S) strain and were reared for generations under 26–28 °C with 65–70% relative humidity in the laboratory of Dr. Toru Miyamoto, Tokyo University of Agriculture.
- 6. Nishiwaki, N.; Sato, K.; Nakagawa, Y.; Miyashita, M.; Miyagawa, H. *J. Pestic. Sci.* **2004**, *29*, 110.
- 7. The synergistic ratio for imidacloprid was 9.2 times to $8.8\,\mathrm{ng/fly}$ in our experiment.
- Nishimura, K.; Tanaka, M.; Iwaya, K.; Kagabu, S. Pestic. Biochem. Physiol. 1998, 62, 172; Nishiwaki, H.; Nakagawa, Y.; Ueno, T.; Kagabu, S.; Nishimura, K. Pest Manag. Sci. 2001, 57, 810; Kiriyama, K.; Itazu, Y.; Kagabu, S.; Nishimura, K. J. Pestic. Sci. 2003, 28, 8; Kagabu, S.; Ishihara, R.; Hieda, Y.; Nishimura, K.; Naruse, Y. J. Agric. Food Chem. 2007, 55, 812.
- 9. Kagabu, S.; Medej, S. Biosci. Biotechnol. Biochem. 1995, 59, 980.
- Kagabu, S. In Chemistry of Crop Protection; Voss, G., Ramos, G., Eds.; Progress and Prospects in Science and Regulation; Wiley-VCH: Weinheim, Germany, 2003; pp 193–212; Jeschke, P.; Nauen, R.. In Comprehensive Molecular Insect Science; Gilbert, L. I., Iatrou, L., Gill, S. S., Eds.; Elsevier: Oxford, UK, 2005; Vol. 5, pp 53–105.
- Unpublished results conducted according to the description in the literature.
 Ohno, I.; Hirata, K.; Ishida, C.; Matsuda, K.; Kagabu, S. Bioorg. Med. Chem. Lett. 2007. 17. 4500.
- Connors, K. A. Binding Constants; Wiley: New York, USA, 1987. Chapters 1–3, 5;
 Hirose, K. In Analytical Methods in Supramolecular Chemistry; Schalley, C. A., Ed.;
 Wiley-VCH: Weinheim, Germany, 2006. Chapter 2.
- 13. Izatt, R. M.; Pawlak, K.; Bradshaw, J. S. Chem. Rev. 1991, 91, 1721.